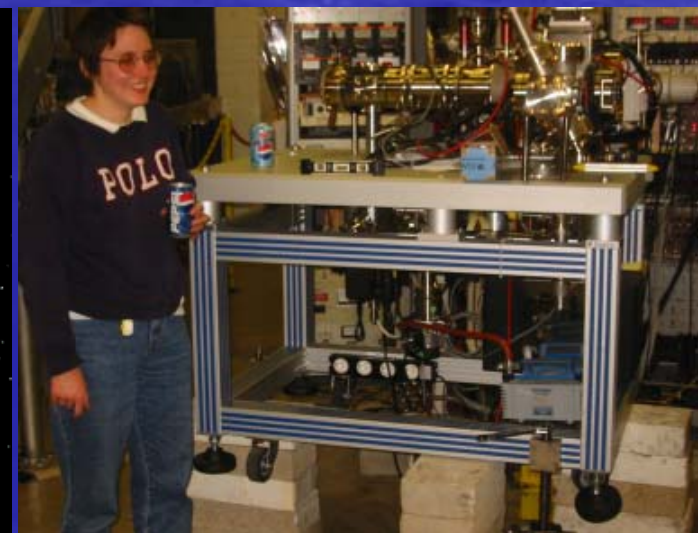


# Soft X-ray spectromicroscopy & its analysis with **aXis2000** .

Adam Hitchcock - BIMR, McMaster University

## GOALS

1. familiarization with Synchrotron spectromicroscopy techniques
2. demonstration of data analysis with aXis2000
3. discussion of potential for your application

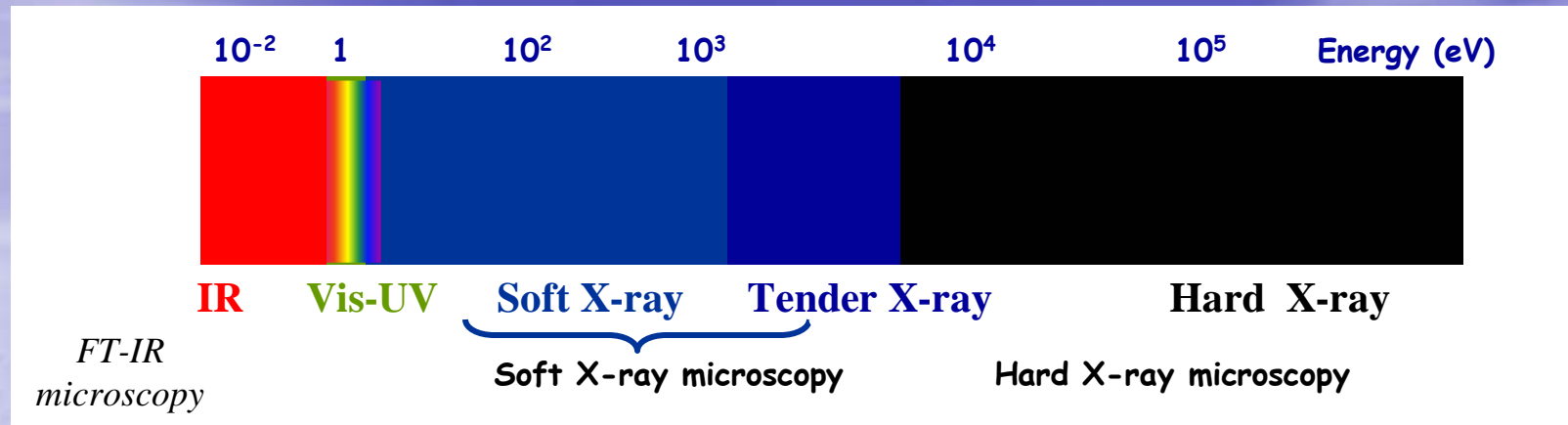


# Outline

- Modes & information from X-ray spectromicroscopy
- Data analysis - example from STXM  
chemical mapping of protein on a phase segregated polymer
- structure of aXis2000 widget
- how can you access the power of aXis2000 ?



# Basic Principles



- \* Use **X-ray absorption contrast** for
  - \* chemically sensitive imaging - "X-ray imaging" **NEXAFS microscopy**
  - \* spatially resolved chemical analysis - "Micro-probe"
- \* Use **penetrating power** of X-rays to study
  - \* wet soft matter (biology, polymers, nano-materials) **Soft X-rays ("water window")**
  - \* fluorescence microprobe **Hard X-rays**
  - \* non-destructive testing; tomography

## References:

- J. Kirz, C. Jacobsen and M. Howells, *Quarterly Review of Biophysics*, **33** (1995) 33
- H. Ade, in *Experimental Methods in the Physical Sciences*, Vol. 32, pp. 225, J.A.R. Samson and D.L. Ederer Ed., Academic Press, 1998
- A.P. Hitchcock, *American Laboratory*, **33** (2001) 30; *J. El. spec.* **144** (2005) 259.
- H. Ade and S.G. Urquhart, in "*Chemical Applications of Synchrotron Radiation*" T. K. Sham, ed. (World Scientific Publishing, 2002)

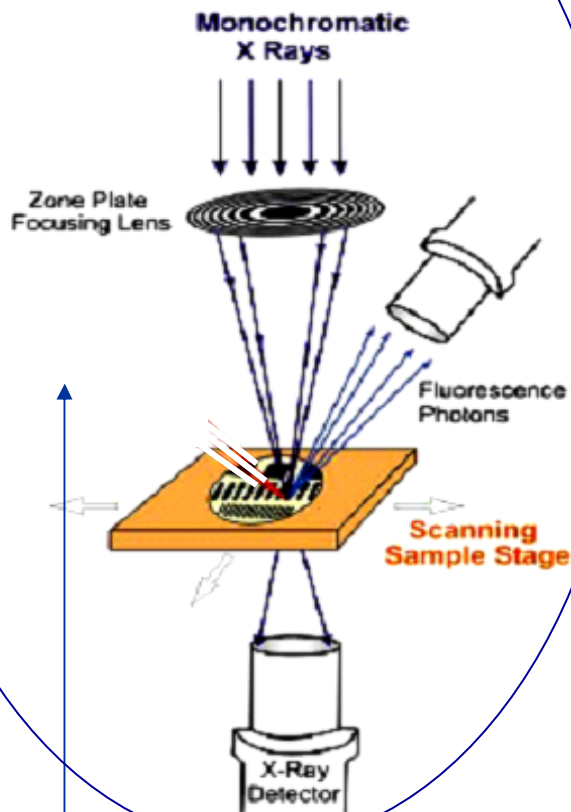
# Overview of techniques

STXM

ALS BL 5.3.2; BL11.0, NSLS X1A, BESSY

TXM now at ALS BL 6.1.2 XM-1, BESSY

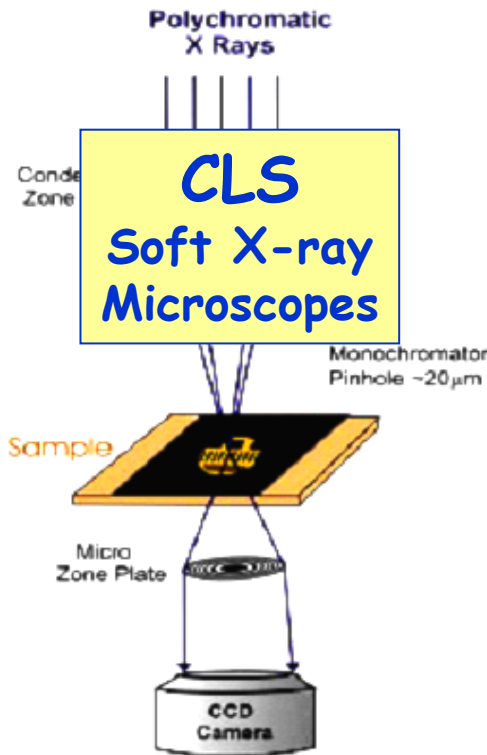
Scanning Transmission X-ray  
Microscopy - STXM



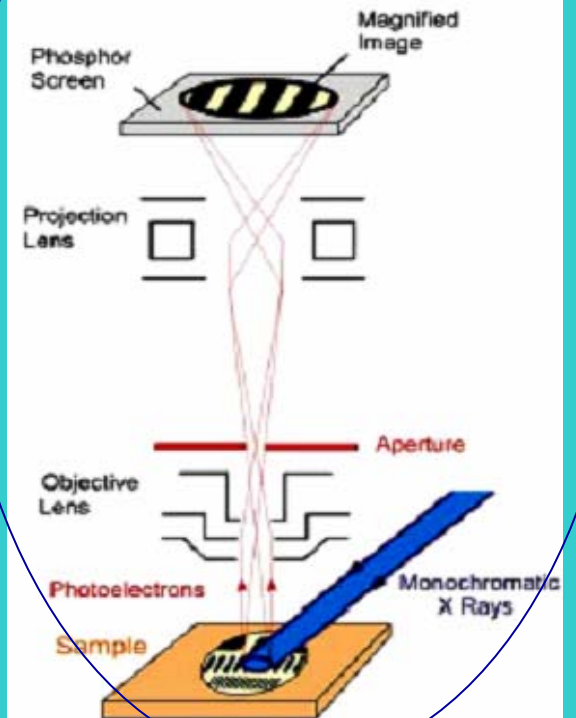
Scanning Photoelectron  
Microscopy - SPEM

SPEM now at ALS, Trieste, Taiwan, Korea..

Transmission X-ray  
Microscopy TXM



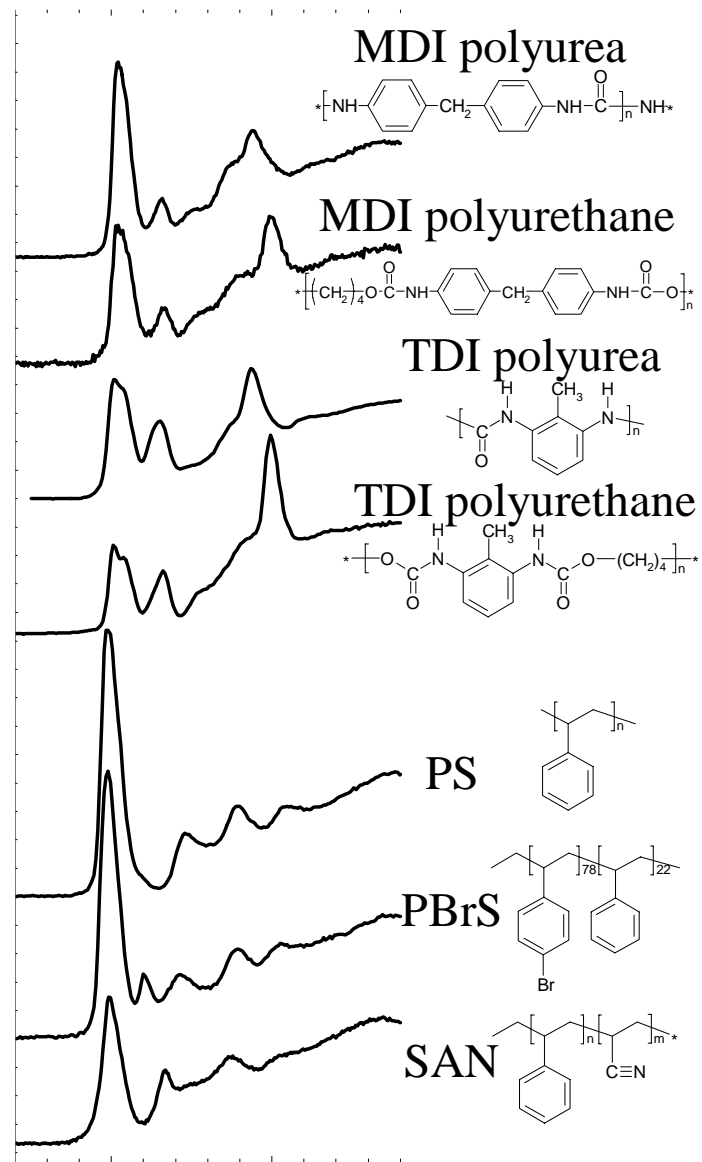
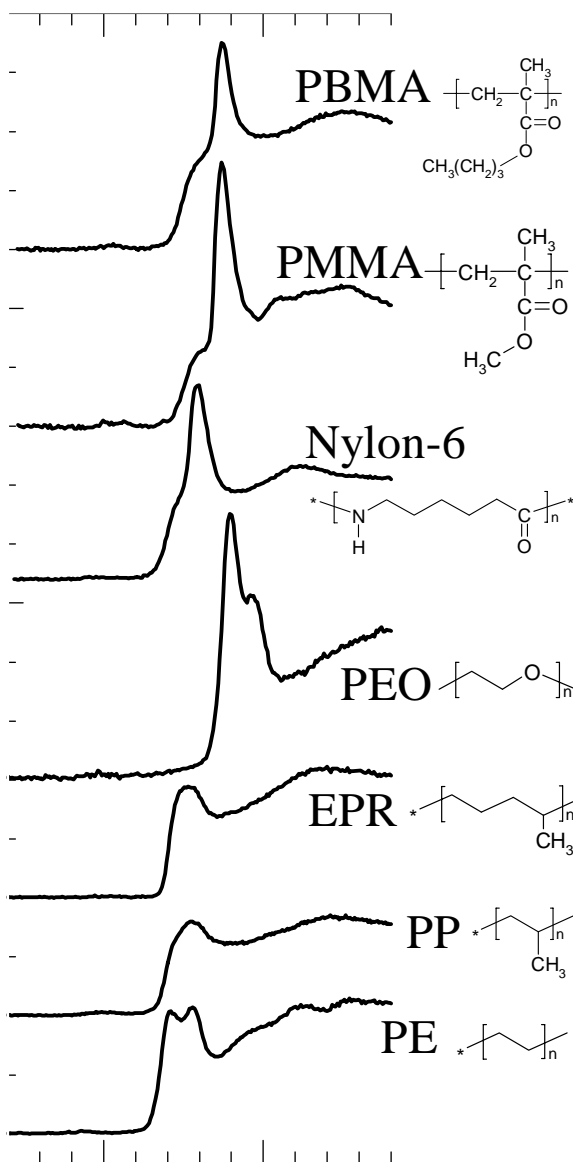
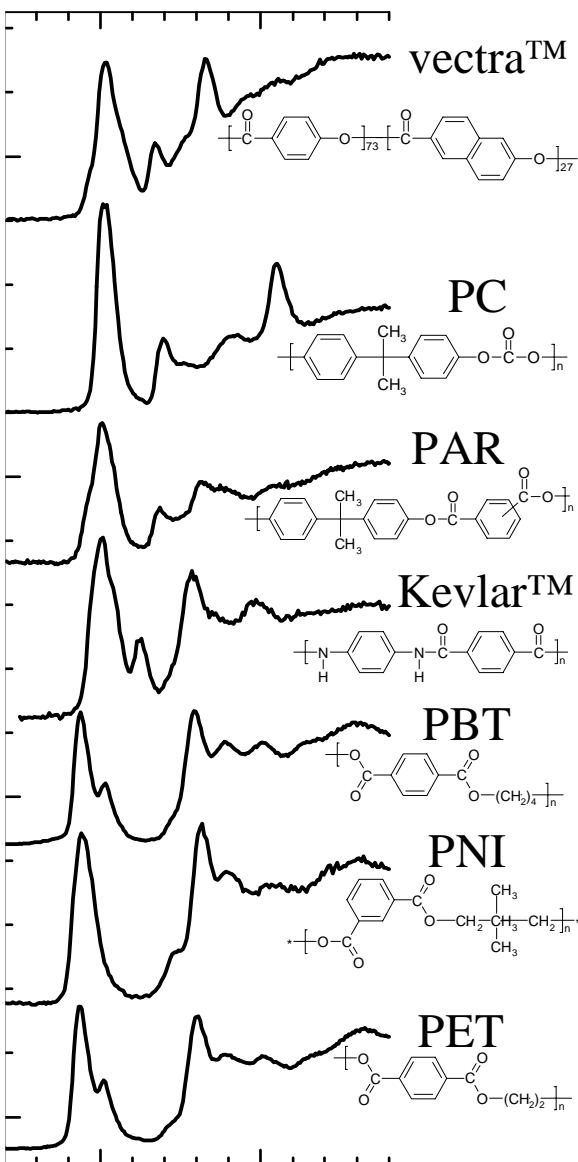
X-ray PhotoEmission Electron  
Microscopy - X-PEEM



X-PEEM now at ALS BL 7.3.1, SLS, Trieste, SRC

## Sensitivity of Polymer NEXAFS Spectroscopy

**Ade , Urquhart (1997-99)**  
**(nsls X1A stxm)**



285      290  
Photon Energy / eV

285      290  
Photon Energy / eV

285      290  
Photon Energy / eV

# Electron yield-based soft X-ray microscopies

Primary XAS process produces - **photoelectrons**

Core hole decay produces

- Auger & **secondary electrons**
- photons
- ions
- luminescence photons

All are being developed as detection channels for analytical X-ray microscopy

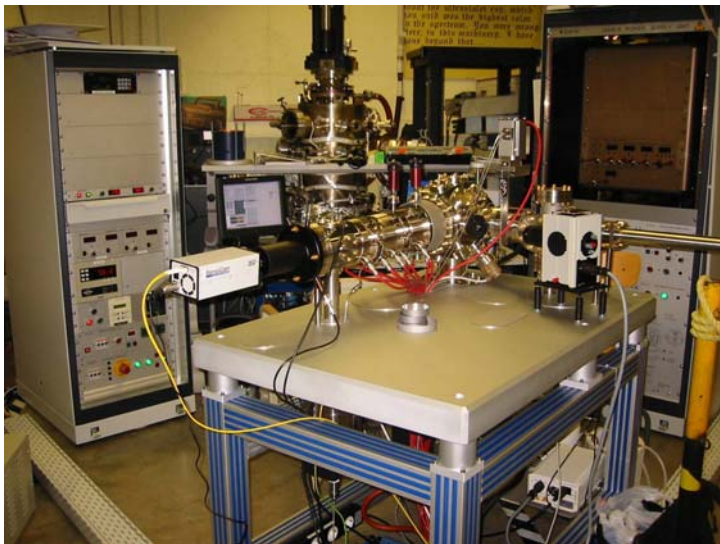
SPEM – Scanning PhotoElectron Microscopy

PEEM - Photo-Emission Electron Microscopy

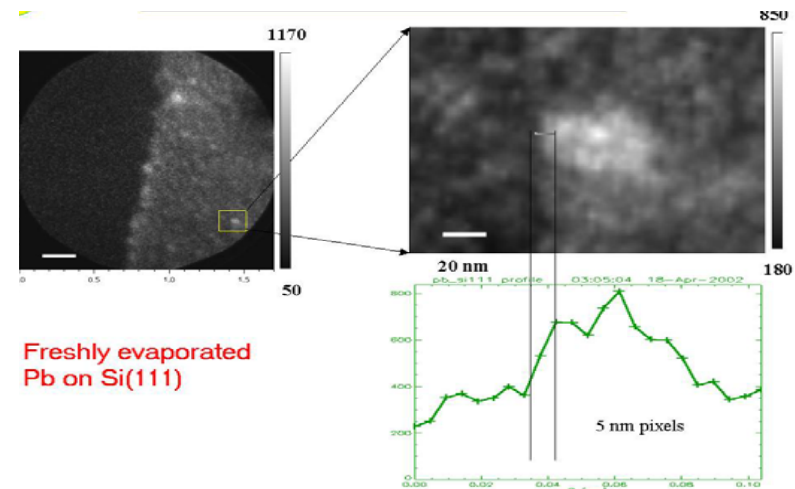
Elettra, Pohang Light Source  
**ALS BL 7.0.1**

*Commercially available*

\* most SR facilities ; BESSY -**SMART**  
**ALS BL 7.3.1 [ PEEM2]**



**CaPeRS** – Canada's Elmitec PEEM  
now at CLS

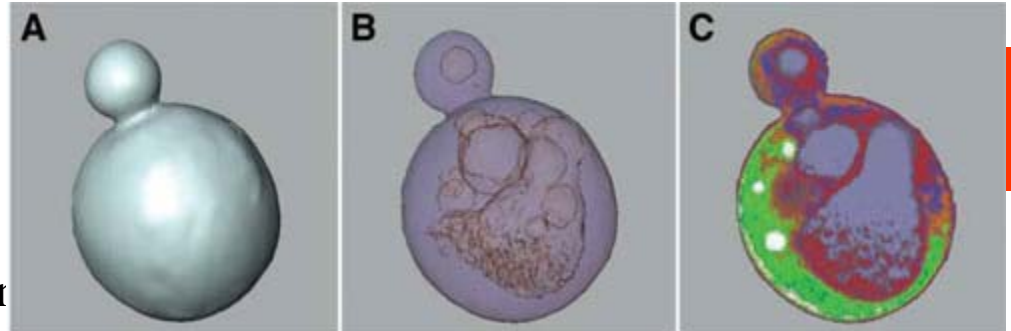
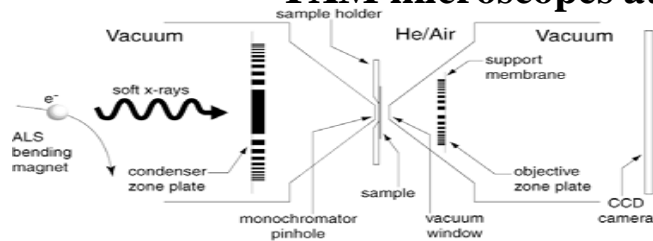


But only ~50 nm with synchrotron light due to e- distribution & chromatic aberration

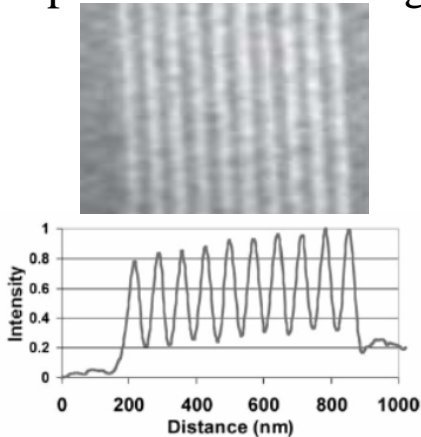


# TXM – biological applications

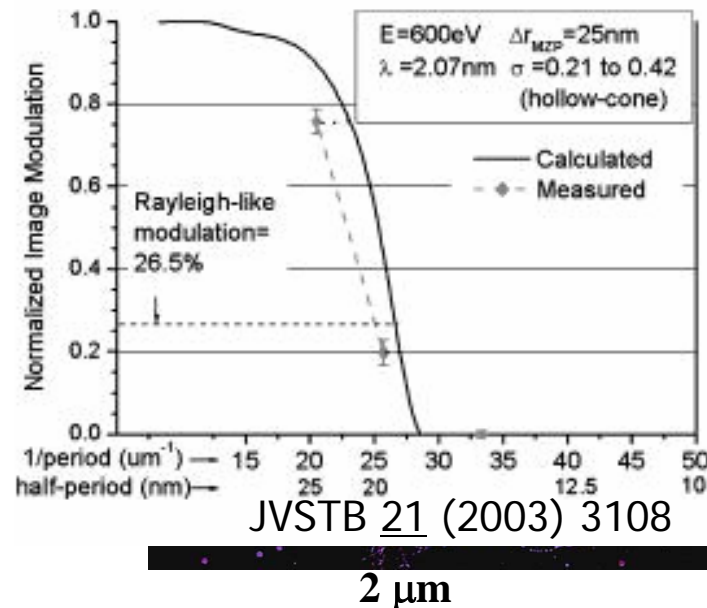
- first demonstrated Gunter Schmäl (Gottenberg) – BESSY (~1986)
- TXM microscopes at ALS (XM-1), Bessy, Aarhus, Elettra



- wet cells: biomaterials & biological imaging
- cryo imaging
- tomography
- XMCD imaging
- phase contrast imaging

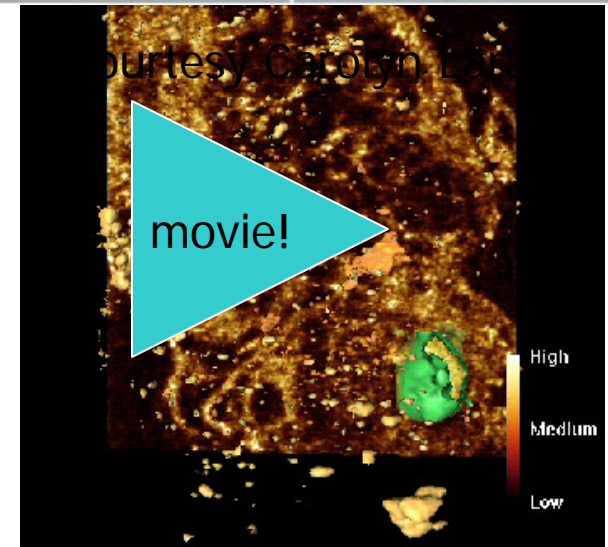


Highest spatial resolution  
22 nm diffraction limit achieved  
 test sample: 15 nm lines 4:1 spacing



monoclonal antibody / Au-Ag-labeling of  
 cytoskeleton proteins

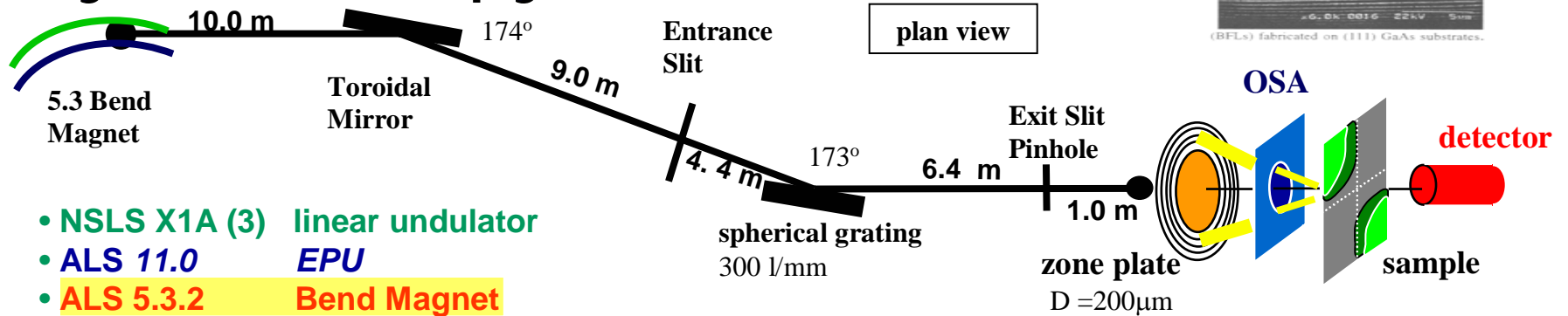
*Larabell (UC Davis) ALS 1999*



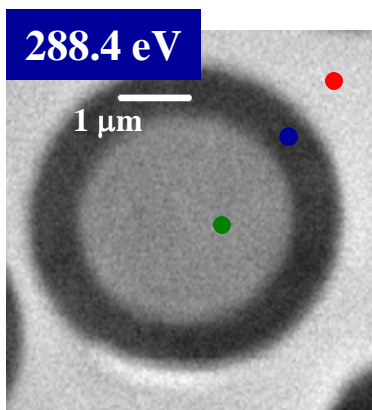
*Gerd Schneider, Carolyn Larabell  
 ALS XM-1 2003*

Angle-scan tomography  
 (cf. Attwood)

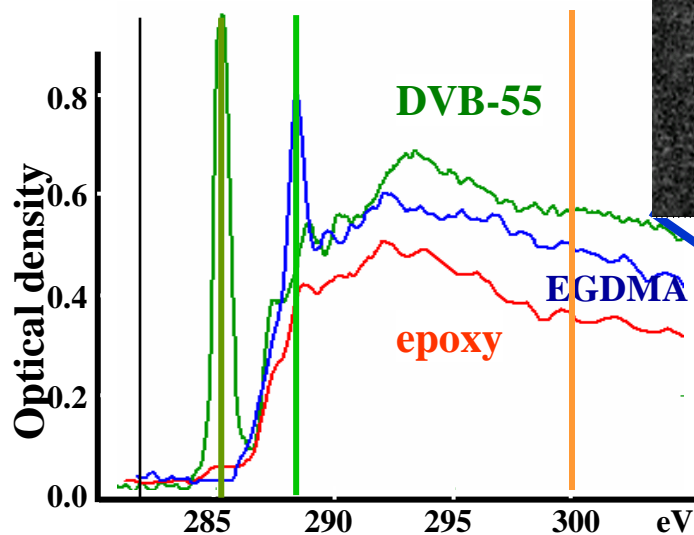
# Scanning Transmission X-ray Microscopy (STXM)



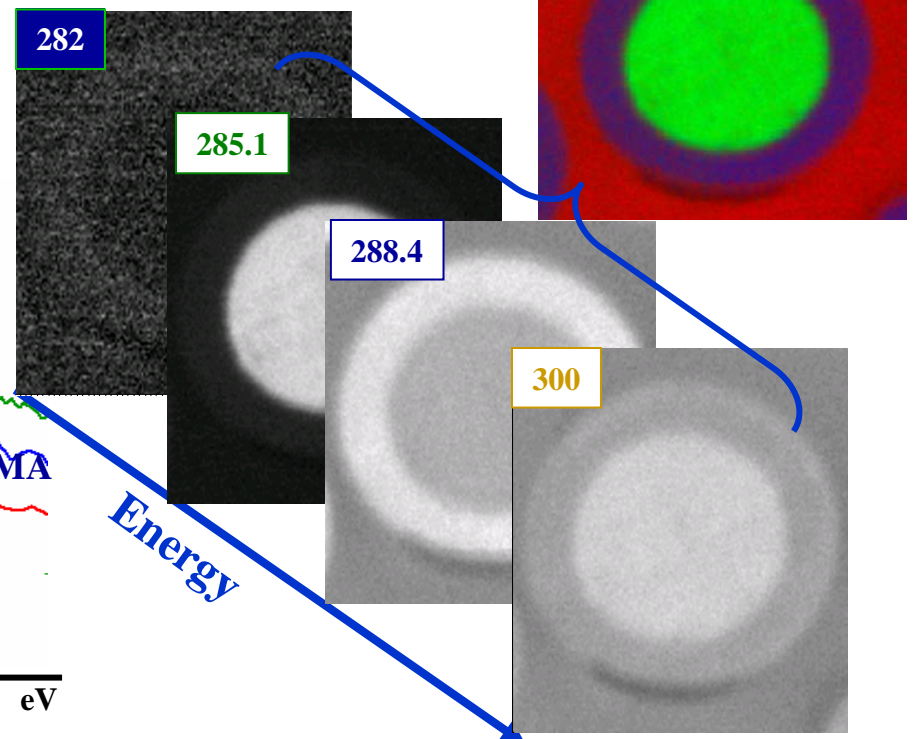
**Images**  
transmitted  $I(x,y)$



**Spectra  $\ln(I/I_0)$**   
{point, line, image}

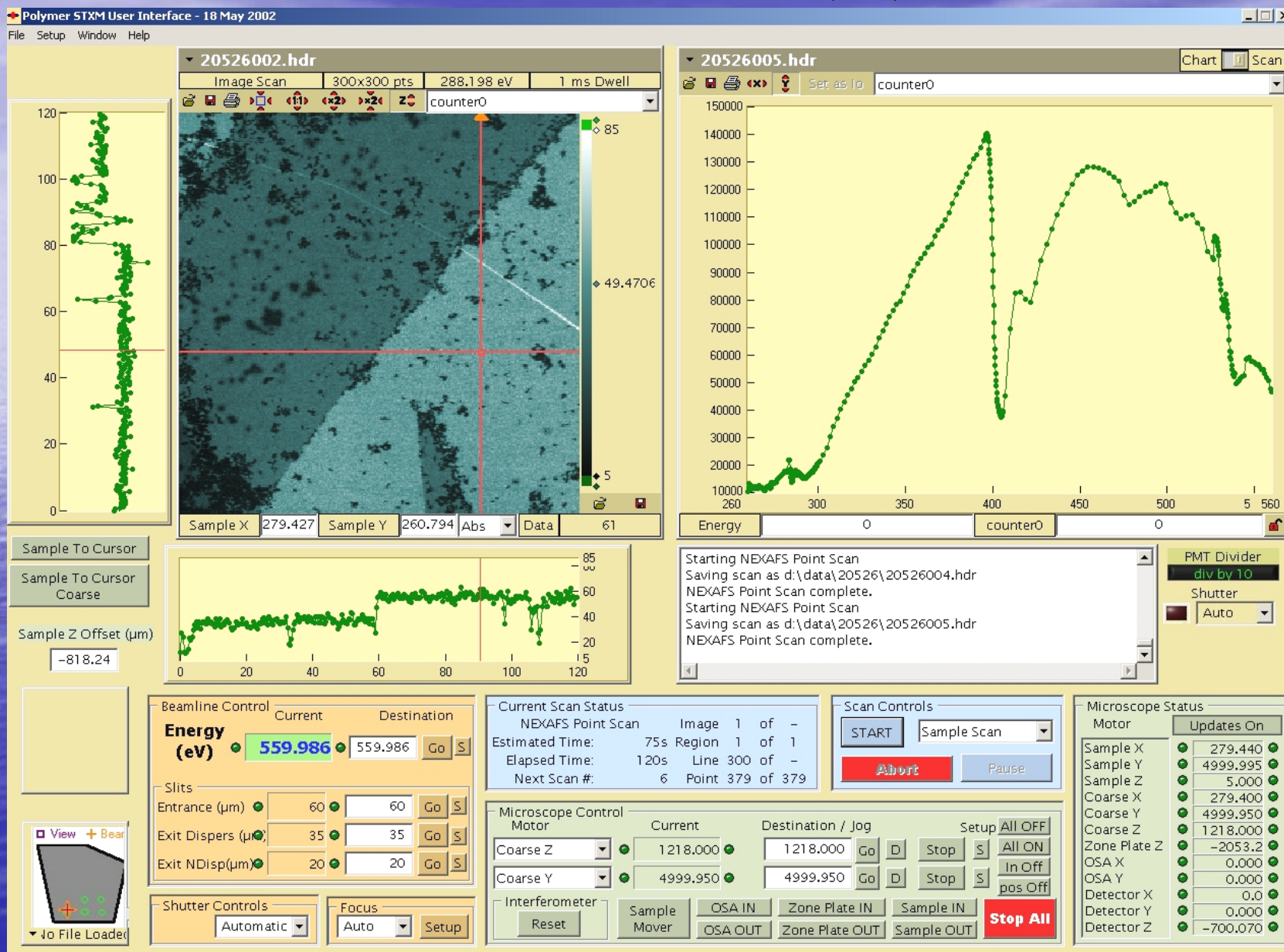


**Image sequences (OD format)**



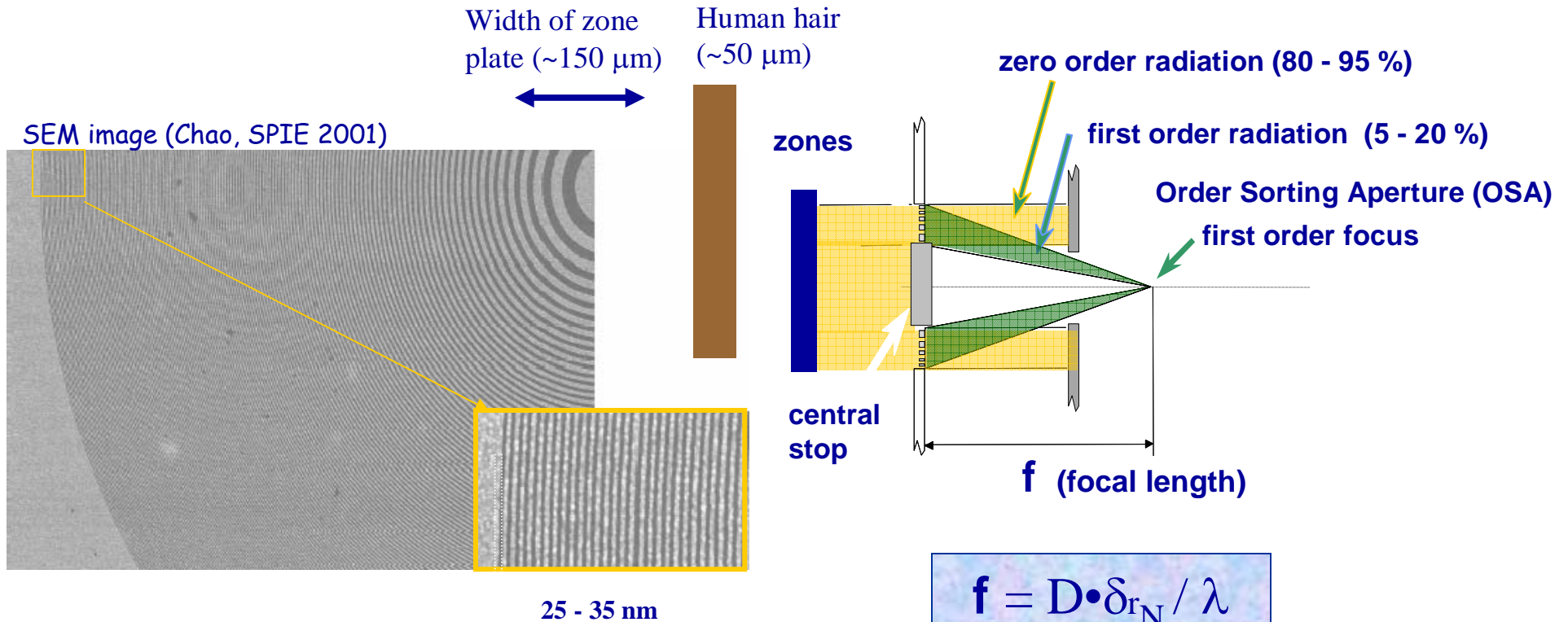


# STXM\_control - ALS (2), CLS, SLS



# Fresnel zone plates: diffractive focusing

A.G. Michette, Optical Systems for soft X-rays, Plenum Press, 1986



**Typical Values** (for current ZP in stxm532)

$\lambda$  (photon wavelength) 1 to 6 nanometers ( $\sim 1240/E$ )

$D$  (ZP diameter) = 155 microns

$\delta r_N$  (outer zone width) = 35 nanometers

Number of zones  $\sim 1000$

Central stop diameter = 80 microns

OSA diameter = 55 microns

$$f = D \cdot \delta r_N / \lambda$$

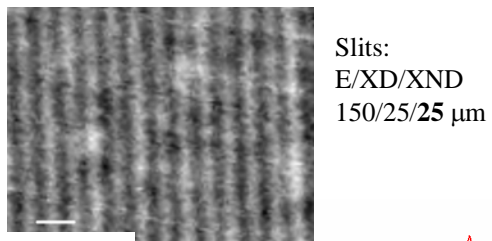
0.9 -1.2 mm in C 1s region

Spatial resolution  
(diffraction limited)

$$\Delta r = 1.22 \cdot \delta r_N$$

# 5.3.2 STXM Performance

- Diffraction limited spatial resolution (40 nm)
- 50 meV spectral resolution
- 50 nm CHEMICAL resolution via interferometry



Slits:  
E/XD/XND  
150/25/25  $\mu\text{m}$

100 nm

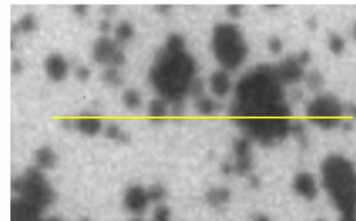
~10% contrast

26 nm

Nov 2002

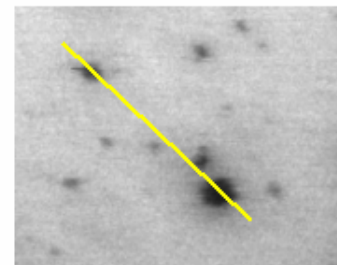


ALS BL 5.3.2 (Aug-01)

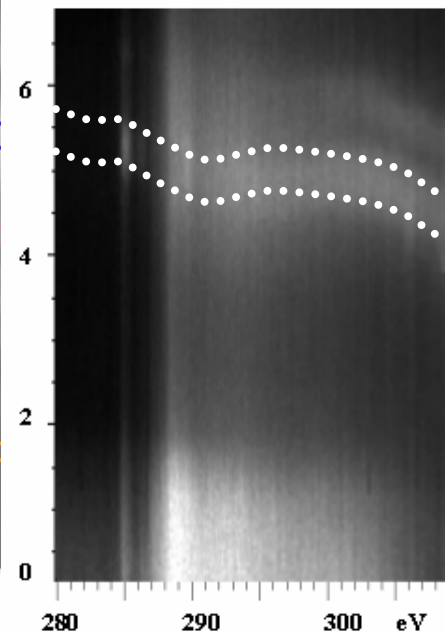
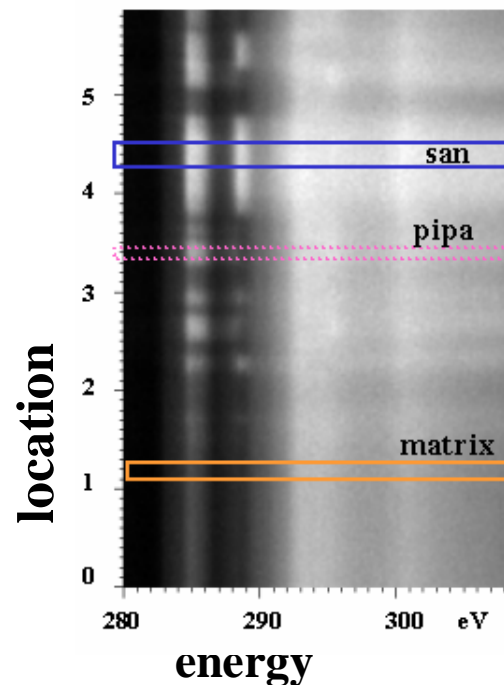


500 nm

ALS BL 7.0 (Dec-98)



1000 nm



Advanced Light Source

Tony Warwick

Mike Kirschner

Keith Franck



NC STATE UNIVERSITY

David Kilcoyne

Harald Ade (NCSU)



Tolek Tyliszczak

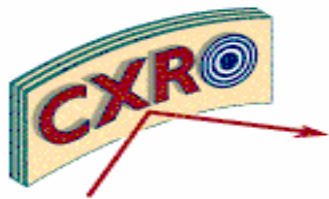
Adam Hitchcock

Peter Hitchcock



Living.  
Improved daily.





# STXM-11: state-of-art performance

December 2003 - BREAKTHROUGH in ZP technology !

## 25 nm diffraction limited zone plates

Recent advances in CXRO zone plate fabrication has resulted in new STXM zone plates with significant improvements in

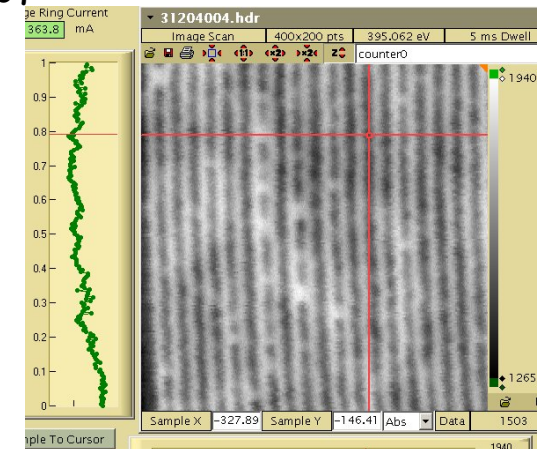
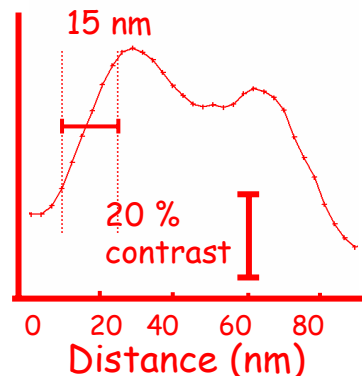
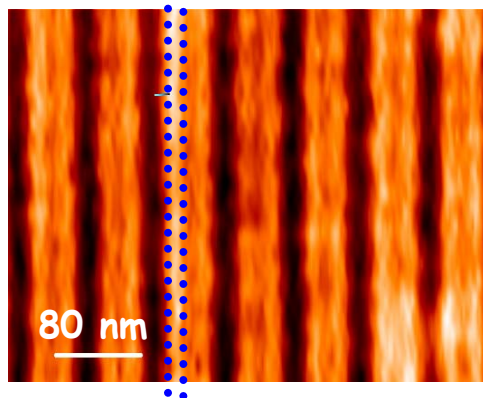
- spatial resolution
  - due to narrower outer most zones (25 nm instead of 35 nm)
- performance at high photon energy
  - due to higher aspect ratio (7:1 instead of 3:1)

The performance of the interferometrically controlled STXM - in particular its thermal and temporal stability, as well as precision of tracking over variable photon energy - has been found sufficient to take advantage of the improved zone plate performance.



25 nm zone plate as test

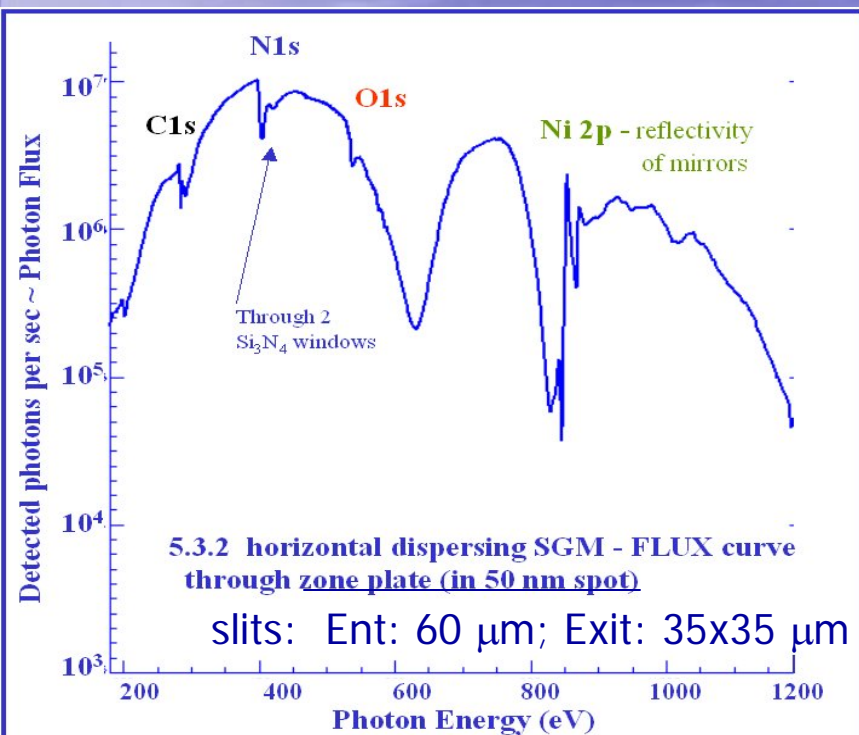
## 25 nm 1:1 lines as test object



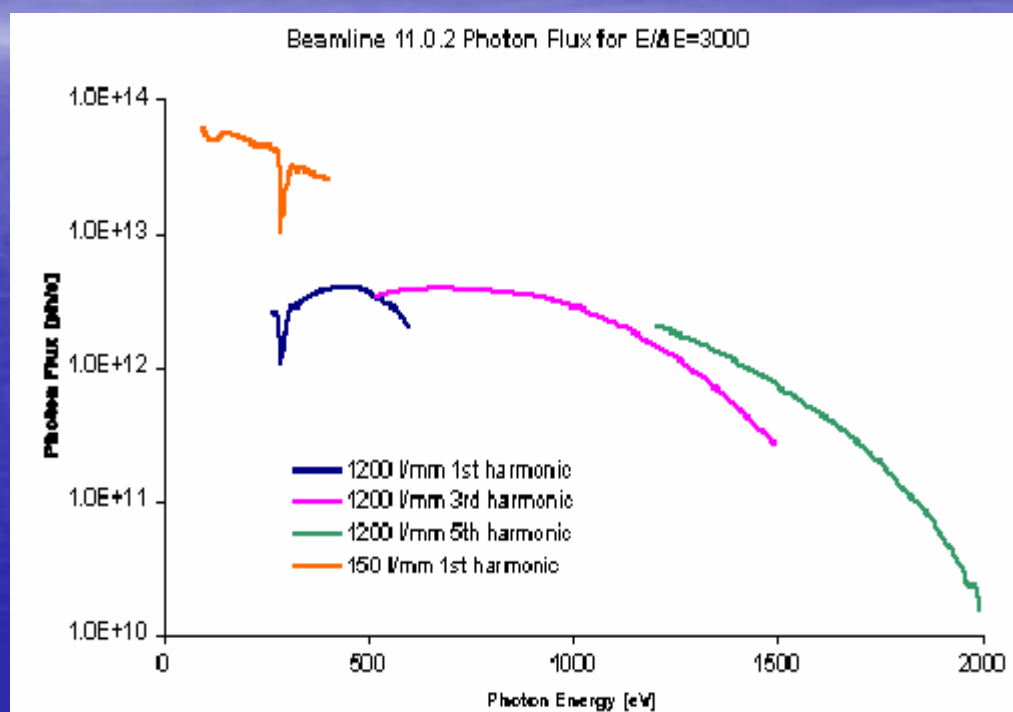
**STXM532** - can mount 25 nm ZPs  
 - usually 35 nm ZPs (intensity)

# ALS STXMs: Energy range, flux

BM STXM 5.3.2



Undulator STXM 11.0.2



with 90/60/60 slits it is easy to get  
>  $10^8$  ph/s in ~60 nm spot on  
sample at 390 eV

exit slits are typically 5-30  $\mu\text{m}$   
>  $10^9$  ph/s ~40 nm spot on  
sample at 390 eV

# Spectromicroscopy at the CLS

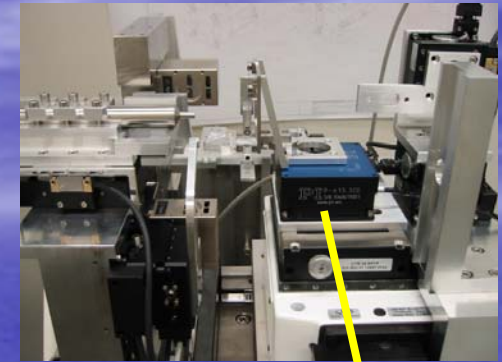
**Insertion device:** Elliptically Polarized Undulator (EPU)

**Monochromator:** Plane Grating, no entrance slit (modified SX-700)

**Energy range:** 250 - 1900 eV

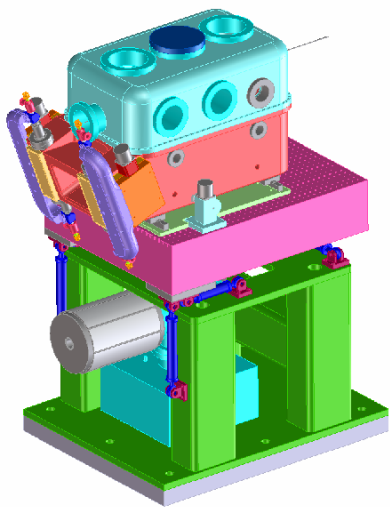
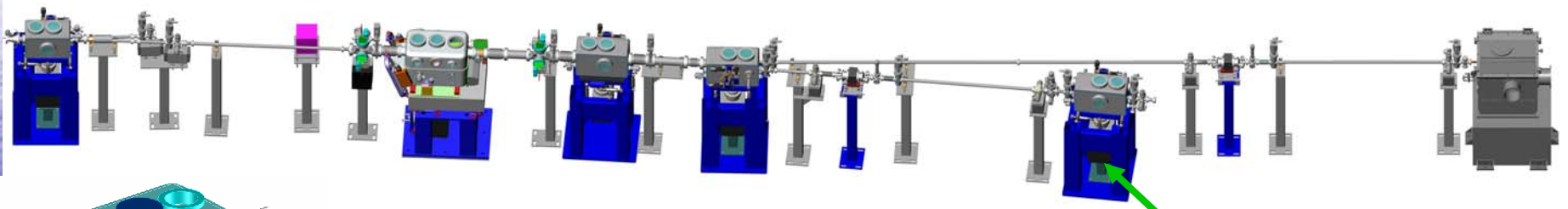
**Resolving power:** 5000

**Flux:** **PEEM:**  $10^{11}$  -  $10^{12}$  photon/s in 20 micron spot  
**STXM:**  $10^8$  photon/s in 50 nm spot



STXM: modified 5.3.2 design

*ALS assistance: monochromator & mirror holders similar to BL 11.0.1*



## X-PEEM (Stephen Urquhart)

- operated Apr02-Mar05 at SRC (Madison, WI)
- now at CLS
- to run on SGM summer 2005



see talk and poster by Kaznatcheev (MSC)

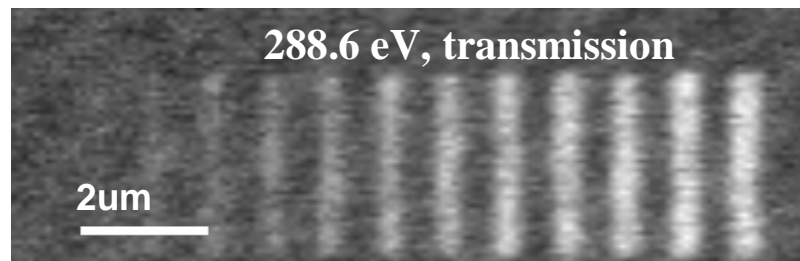
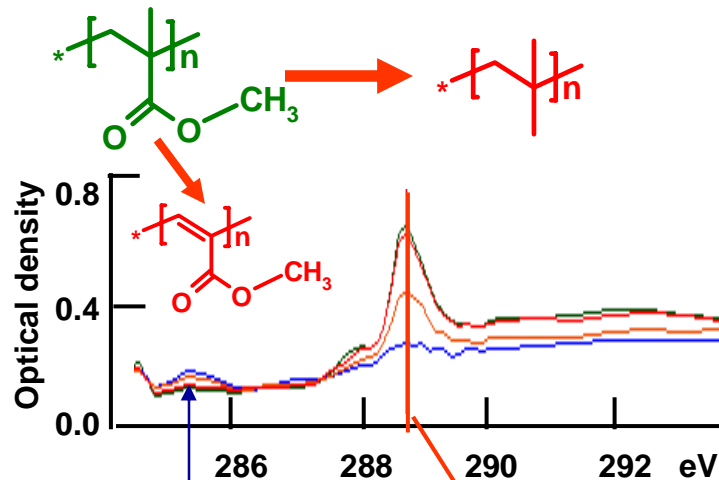


# Radiation damage in STXM: PMMA

\* 4  $\mu\text{m}$  line pairs; spaced at 1  $\mu\text{m}$

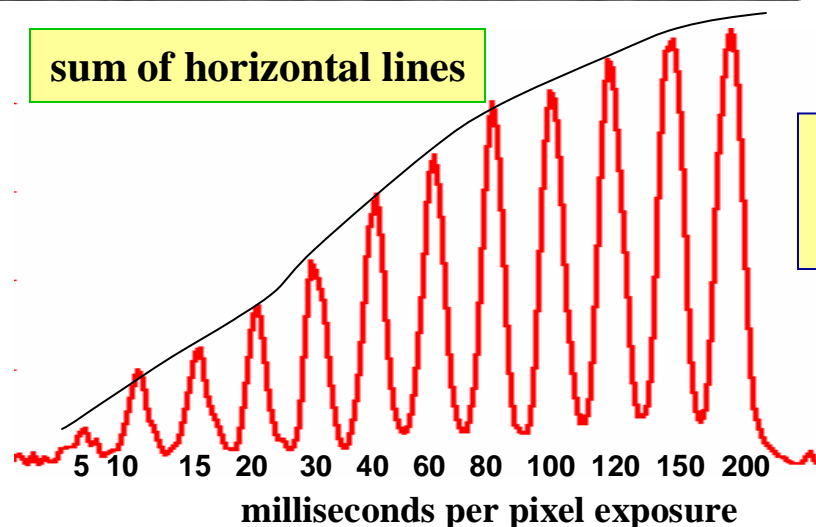
\*\* Exposed in linescan mode at 300 eV;  
~2 MHz in detector

PMMA = polymethylmethacrylate



stxm 5.3.2

sum of horizontal lines



Exposure to  
damage 1/e  
~50 ms !!!

## Damage control measures in STXM 5.3.2

### a) Hardware

- 1 msec in-vacuum piezo shutter
- Closed between successive scan lines
- fast scanning: 0.2 - 1.0 msec/pixel dwell

### b) Acquisition strategy

- Defocused beam (if suitable) – point, line, image modes
- Multi-region acquisition
- Short stacks / SVD

# STXM is optimal for quantitative chemical analysis of soft matter

- ✓ **High Spatial Resolution**

Zone plate properties determine resolution. Typically ~30-40 nm

- ✓ **High Spectral Resolution = high chemical resolution**

All instruments achieve ~ natural line width (0.1 eV in C 1s)

- ✓ **Quantitative compositional analysis**

Beer's Law response – Absorbance (OD) proportional to concentration in column / pixel

- ✓ **Adaptable to many environments**

Fully solvated systems – water window

Magnetic fields

Vacuum – surface analysis

- ✓ **Significantly lower radiation damage** than TEM-EELS . . .

Quantitative comparison indicates  $10^2 - 10^3$  advantage on basis of information / unit damage

PET – Rightor et al J. Phys. Chem. B 101 (1997) 1950

see poster by Wang (MSC)

. . . and TXM     (*In STXM, the inefficient Zone Plate optic is BEFORE the sample*)

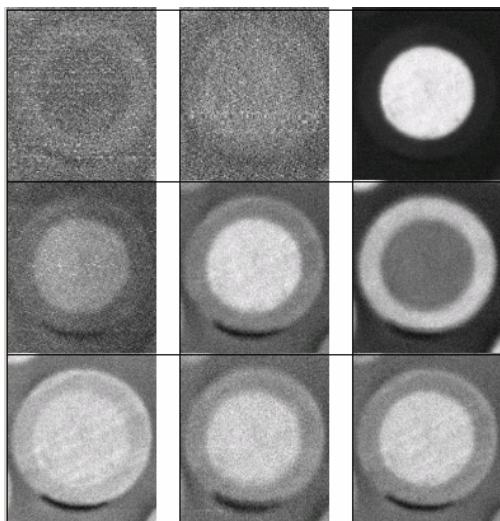
# Quantitative Chemical Mapping

*From pixel-by-pixel fits to reference spectra*

*Implemented in aXis2000*

Core shell particles: (with Stöver)  
see *Macromolecules* **34** (2001) 4424

## 1. Record image sequence (stack)



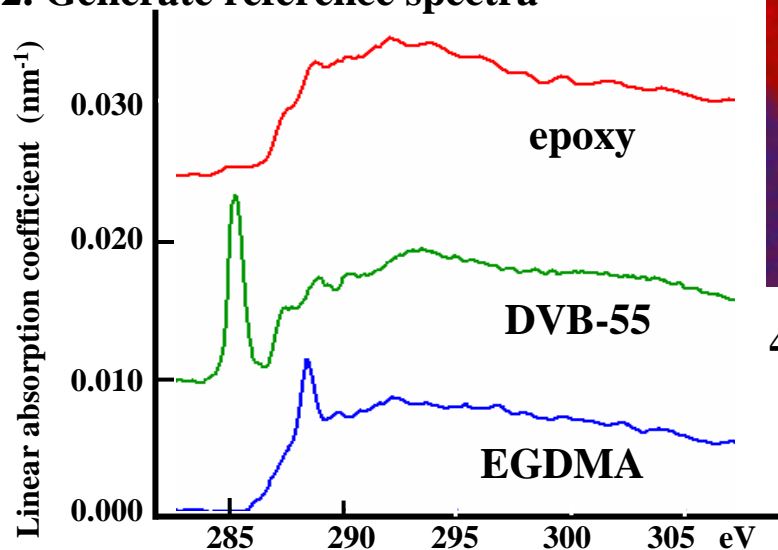
## 3. Generate component maps

$$OD(j,k) = \sum_i a_i(j,k) * (\text{reference})_i$$

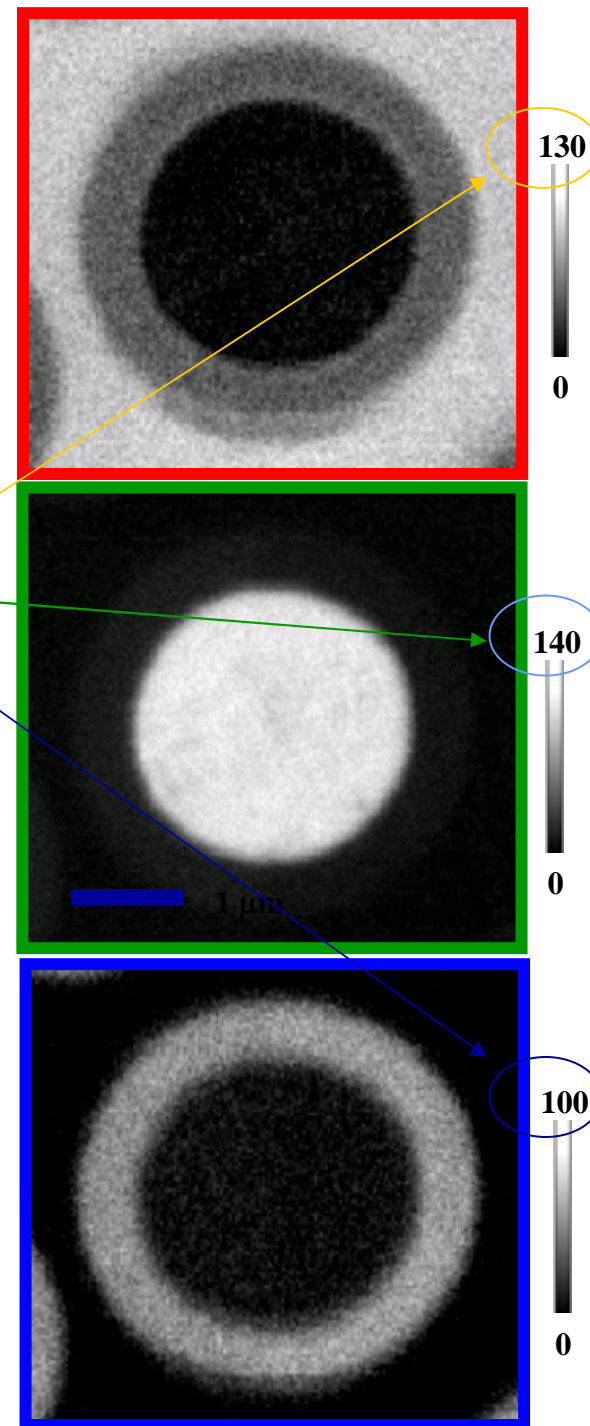
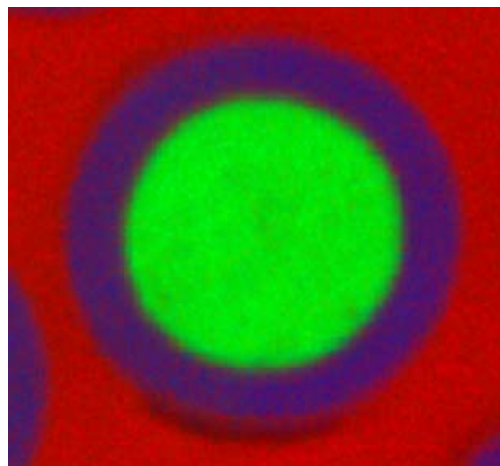
with  $a_i(j,k) = \text{THICKNESS (nm)}$  at  $(j,k)$

when reference spectra are absolute ( $\text{nm}^{-1}$ )

## 2. Generate reference spectra



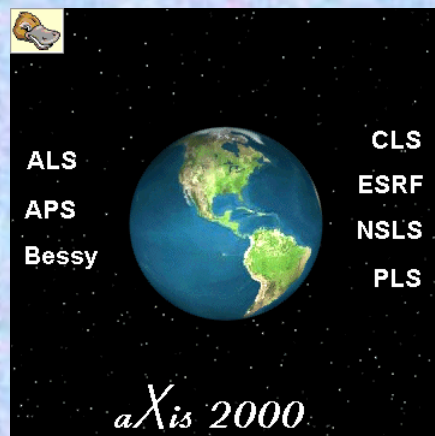
## 4. Generate RGB-composite component “image”





# Analysis software for soft X-ray spectromicroscopy

**aXis2000**



(<http://unicon.mcmaster.ca/aXis2000.html>)

**IDL VM 6.0**  
The IDL Virtual Machine™  
Distribution Platform for IDL Applications

click to continue

Upgrade to a development version of IDL and:

- interactively explore your data in the IDL environment
- develop cross-platform applications for distribution
- test custom data analysis algorithms

Find out more at [www.rsinc.com/IDL](http://www.rsinc.com/IDL)

©2003 Research Systems Inc. RSI Research Systems Inc.

aXis2000 is  
free for non-  
commercial use

5.3.2 STXM can be viewed and even run  
remotely (with training & permission) over the net  
(‘Fedex’ synchrotron microscopy)

## Features of the aXis2000 widget

**Y lineout at X-position of cursor**

**First-row pull-down menus**

**Thumbnails**  
• Click to select a buffer

**Axis Messages, Hints and log**

**Second row, single command**

**X lineout at Y-position of cursor**

**Color-bar for Images**

**10 Data Buffer List**  
• Click to select  
• Use utilities-change label to change label  
• Buffer 0 = modified data  
• Slider for long labels

**Lineouts, symbols & scale bar options**

**Gamma for Images**

**X, Y, (Z) limits for Images & spectra (display & control)**

**Cursors**  
(X,Y,Z) – at cursor  
(dX,dY,dZ) – change over line (images) or between cursors (spectra)  
dR – distance along line (images only)

**Main Image**  
• Displays currently selected image or selected spectrum (or group of spectra, if Spectra-Overplot used)  
• Size of aXis2000 display can be adjusted (0.5 to 2.0) of a nominal size (360x360 pixels in Main Image) by size parameter in axis.ini  
**Mouse** (if Z-lines is selected)  
• **First** click – cursor and lineout; arms the line generator  
• **Second** click – draws and documents line (image) ;  
- reports difference in cursors (spectra)  
• **Third** click – clears line and cursor information

# BREAK TIME !

